Original Article
LncRNA CRNDE promotes hepatic carcinoma cell proliferation, migration and invasion by suppressing miR-384

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Received May 17, 2016; Accepted June 16, 2016; Epub October 1, 2016; Published October 15, 2016

Abstract: Studies have found that colorectal neoplasia differentially expressed (CRNDE) is related to cancer development. Herein, we found that the expression of CRNDE was increased in human hepatic carcinoma (HCC) tissues and cell lines. The ROC curve analysis illustrated CRNDE has a significant diagnostic value for HCC. At the same time, CRNDE promotes HCC cell proliferation, migration, and invasion in vitro. Quantitative real-time polymerase chain reaction (PCR) demonstrated that miR-384 was significantly downregulated in HCC tissues. Moreover, we indicated CRNDE negatively regulated miR-384 expression in HCC. In addition, we found that CRNDE accelerated the expression levels of NF-κB and p-AKT though inhibition of miR-384. Overall, these results suggested that CRNDE-miR-384 axis might be a promising therapeutic target for the treatment of HCC.

Keywords: IncRNA-CRNDE, miR-384, hepatocellular carcinoma, migration and invasion

Introduction

Hepatocellular carcinoma (HCC) is one of the most common tumors to impact human health, accounting for more than 5% of all cancers in the world [1, 2]. It has been reported that HCC has become the second leading cause of cancer mortality in China [3]. Although surgical treatment is the most common and efficient method for treating HCC, the recurrence and metastasis rates are high. According to earlier reports, the 5-year survival rate after operation ranges between 36-50% [4]. HCC cell migration and invasion are very complex processes, including changes in multi-gene signatures. The early prevention of migration and invasion has become a primary topic in HCC research. Therefore, the molecular mechanisms of potential metastasis in HCC and the effective therapeutic targets require urgent exploration.

Long non-coding RNAs (lncRNAs), a new sort of non-coding RNA longer than 200 nucleotides, have no protein-coding capacity [5]. Numerous studies have indicated that lncRNAs regulate gene expressions through the processes of transcription regulation, post-transcription regulation, genomic imprinting, and chromatin modification [6, 7]. There is already growing evidence that lncRNAs play major roles, not only in normal development but also in tumor genesis [8, 9]. However, the biological function and mechanism of IncRNA-CRNDE are not clear in HCC. In this research, we measured the expression level of IncRNA-CRNDE in human hepatic carcinoma cell lines and tissues. We also demonstrated that IncRNA-CRNDE promoted HCC cell proliferation, migration, and invasion in vitro.

MicroRNAs (miRNAs), a class of endogenous non-coding RNA with the length of 20-25 nucleotides, can regulate the expression of massive target genes by targeting the homologous sequences of messenger RNAs (mRNAs) with the 3'-untranslated region (3'-UTR) to promote RNA degradation [10, 11]. MiRNAs play key roles in the regulation of cell development process including proliferation, differentiation, metastasis, and apoptosis [12, 13]. However, the mechanism of miR-384 action is not clear in HCC.
LncRNA-CRNDE regulates miR-384 in HCC

In this study, our results revealed a novel mechanism of lncRNA-CRNDE in tumorigenesis of HCC. We surmised that lncRNA-CRNDE can directly interact with miR-384 to promote HCC cell proliferation, migration, and invasion in vitro. Therefore, we speculated that the lncRNA-CRNDE, as a key regulator of gene expression, might be a promising therapeutic target for the treatment of hepatic carcinoma.

Materials and methods

Patients and clinical specimens

HCC tissues and matched adjacent noncancerous tissues samples were collected and written, informed consent was provided by patients in the Department of Hepatobiliary Surgery, Affiliated Hospital of Guizhou Medical University. According to World Health Organization (WHO), the HCC histological diagnosis was confirmed.

Cell culture

Seven HCC cell lines (MHCC97H, Hep3B, MHCC97L, BEL-7402, QGY-7703, HCCC9810, and HuH7), immortalized normal liver epithelial cell lines (THLE3), and human embryonic kidney 293T (HEK293T) cell lines were purchased from the Type Culture Collection of the Chinese Academy of Sciences, Shanghai, China. MHCC97H, Hep3B, MHCC97L, BEL-7402, QGY-7703, HCCC9810, HuH7, and HEK293T were cultured in Dulbecco’s modified Eagle’s medium (Invitrogen, Carlsbad, CA) containing 10% fetal bovine serum (FBS, Invitrogen), 100 U/ml penicillin, and 1 μg/ml streptomycin (Invitrogen). THLE3 cells were cultured in bronchial epithelial growth medium (Clonetics Corporation, Walkersville, MD), including 5 ng/ml epithelial growth factor (EGF), 70 ng/ml phosphoethanolamine, and 10% FBS. All cells were cultured at 37°C with 5% CO₂.

Lentiviral vector construction

The vectors carrying GFP, lncRNA-CRNDE, Luc, and shRNA-CRNDE were constructed. The packaging vectors (pCMV-VSVG, pMDLg/pRRE and pRSV-REV) and the lentivirus vectors were co-transfected in HEK293T cells, and then lentiviruses were concentrated and identified.

Transfection

MHCC97H cells were transfected with Lenti-GFP and Lenti-lncRNA-CRNDE. HuH7 cells were transfected with Lenti-shCRNDE, and Lenti-shLuc using 8 μg/mL polybrene (Sigma). G418 (Life Technologies, 0.8 mg/mL) was used to screen and establish the stable expression of cell lines. According to the manufacturer’s protocol, MHCC97H and HuH7 (2 × 10⁵ cells/well) were transfected with 200 μl of mature miR-384 mimic (100 nM), mock, or inhibitor with Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) for 72 hrs.

Reverse transcription and quantitative real-time reverse transcription PCR (qRT-PCR)

Total RNA was extracted using the TRIzol reagent (Invitrogen, CA, USA). Corresponding cDNA was synthesized using random primers and a RevertAid First Strand cDNA Synthesis kit (Thermo Fisher). SYBR-Green PCR Master Mix kit (Takara, Japan) and ABI 7500 Real-Time PCR System (Applied Biosystems) were used to detect the mRNA expression levels of related genes. GAPDH and U6 were chosen as internal loading controls. The sequences of GAPDH primers were 5'-TGTTAAATCGGAATTTTAA-3' (sense) and 5'-TGTTACAGGCTTTATGAA-3' (antisense); The primer sequences for U6 are: 5'-CTCGCTTCGGCAGCACA-3' (sense) and 5'-GCCGCTACGACCA-AATC-3' (antisense); The primer sequences of miR-384 primers were 5'-GTCAGCGCCATCTTCTTTG-3' (antisense); The sequences of lncRNA-CRNDE primers were 5'-TGTTAAATCGGAATTTTAA-3' (sense) and 5'-TGTTACAGGCTTTATGAA-3' (antisense); The sequences of NF-κB primers were 5'-AGTGTGGAGGCTGCTGCGAATG-3' (sense) and 5'-TGGCCTTTCAAGCTGGAGTTG-3' (antisense); The sequences of NF-κB primers were 5'-AGTGTGGAGGCTGCTGCGAATG-3' (sense) and 5'-TGGCCTTTCAAGCTGGAGTTG-3' (antisense); The sequences of shRNAs were 5'-GTTGTCAGTCTGGTGCGATG-3' (antisense); All results were repeated three times. All results are shown as the mean ± SD of three independent experiments.

Western blot analysis

Cells were harvested in a lysis buffer containing a protease inhibitor cocktail (Roche Applied Science). The concentrations of total proteins were measured using a BCA protein assay kit (Thermo Fisher Scientific). The objective proteins were separated with 8% SDS polyacrylamide gel and transferred to polyvinylidene difluoride (PVDF) membranes (Millipore, Carlebad, CA, USA) and probed with specific antibodies against GAPDH and β-actin. Fluorescent signals were detected using an Odyssey western blotting system (LI-COR, Inc., Lincoln, NE).
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The membrane was incubated with primary antibodies including anti-rabbit-p-AKT (1:1000 dilutions, Santa Cruz, USA), anti-rabbit-NF-kB antibody (1:1000 dilutions, Cell Signaling Technology, Beverly, MA, USA), anti-rabbit-GAPDH antibody (1:4000 dilution, Cell Signaling Technology, Beverly, MA, USA). The enhanced chemiluminescence (ECL) substrate kit (Amersham Biosciences) and the enhanced chemiluminescence detection system (Amersham Biosciences) were used to detect the results.

Cell proliferation

The proliferate ability of HCC cells was measured using 3-(4, 5-dimethylthiazole-2-yl)-2, 5-diphenyl tetrazolium bromide (MTT). Next, 2 × 10^3 cells were seeded in a 96-well plate. At particular points in time, 20 μl MTT solutions (0.5 mg/ml, Sigma) were added to each well for 4 hrs at 37°C. The original culture was removed, and then 100 μl of dimethyl sulfoxide solutions (Sigma) were added to each well. The absorbance was detected by a Tecan plate reader at 490 nm. Each experimental condition was detected in quintuplicate. All experiments were performed in triplicate.

Migration and invasion assays

The migratory and invasive ability of HCC cells were assessed by migration assay with transwell (Corning Life Sciences, Bedford, MA) and the matrigel invasion (BD Biosciences, San Diego, CA, USA) assay, respectively. Cells were seeded to cell culture inserts (Corning Costar Corp) for incubation for 24 hrs at 37°C and then fixed using 4% paraformaldehyde. They were then stained using 0.1% crystal violet solution. The cells above the upper surface were cleared. The number of migratory cells was counted with the microscope.

Dual luciferase reporter assay

StarBase v2.0 (http://starbase.sysu.edu.cn/) was used to predict the binding sites between LncRNA-CRNDE and miR-384. HEK293T cells (5 × 10^4 cells/well) were co-transfected with the constructed plasmids using Lipofectamine 2000 (a renilla plasmid as internal reference) for 48 hrs. According to the manufacturer’s instructions, the luciferase activities were detected by Dual-Luciferase Reporter Assay kit (Promega).

Figure 1. The expression level of IncRNA-CRNDE increases in human HCC cell lines and tissues. A. The expression level of IncRNA-CRNDE was detected by qRT-PCT in THLE3 and HCC cell lines (MHCC97H, Hep3B, MHCC977L, BEL-7402, QGY-7703, HCCC9810, and HuH7), U6 was used as an internal reference, (*P < 0.05, **P < 0.01, ***P < 0.001). B. The qRT-PCR was used to detect the mRNA expression level of IncRNA-CRNDE in 87 pairs of HCC tissues and paired adjacent normal tissues. C. The cut-off score of IncRNA-CRNDE expression was evaluated by a receiver operating characteristic curve (ROC) analysis.
LncRNA-CRNDE regulates miR-384 in HCC

Table 1. The relationship of lncRNA-CRNDE and miR-384 expression levels (ΔCt) with clinicopathological factors in HCC tissues

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>No. of patients (%)</th>
<th>LncRNA-CRNDE</th>
<th>miR-384</th>
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</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Mean ± SD</td>
<td>P value</td>
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<tr>
<td>Total no. of patients</td>
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<td>Age (year)</td>
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<tr>
<td>&gt; 60</td>
<td>38 (43.7)</td>
<td>10.65 ± 2.23</td>
<td>0.09</td>
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<tr>
<td>≤ 60</td>
<td>49 (56.3)</td>
<td>11.87 ± 1.67</td>
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</tr>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>35 (40.2)</td>
<td>11.74 ± 2.12</td>
<td>0.08</td>
</tr>
<tr>
<td>Female</td>
<td>52 (59.8)</td>
<td>11.23 ± 2.32</td>
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<tr>
<td>Lymphatic metastasis</td>
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<td></td>
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</tr>
<tr>
<td>N0</td>
<td>45 (51.7)</td>
<td>11.93 ± 2.03</td>
<td>0.012*</td>
</tr>
<tr>
<td>N1-N2</td>
<td>42 (48.3)</td>
<td>9.65 ± 1.81</td>
<td></td>
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<tr>
<td>Distal metastasis</td>
<td></td>
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<tr>
<td>M0</td>
<td>58 (66.7)</td>
<td>11.58 ± 2.27</td>
<td>0.07</td>
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<tr>
<td>M1</td>
<td>29 (33.3)</td>
<td>10.17 ± 1.94</td>
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<tr>
<td>0 &amp; I &amp; II</td>
<td>49 (56.3)</td>
<td>11.93 ± 2.03</td>
<td>0.012*</td>
</tr>
<tr>
<td>III &amp; IV</td>
<td>38 (43.7)</td>
<td>10.65 ± 1.81</td>
<td></td>
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</tbody>
</table>

*p < 0.05; **p < 0.01.

Statistical analysis

Statistical significance was analyzed by Student's t-test and one-way analysis of variance (ANOVA) using SPSS 15.0 software and GraphPad (GraphPad Prism Software, La Jolla, CA, USA). All results were shown as the means ± SD. P < 0.05 was considered statistically significant.

Results

The expression level of IncRNA-CRNDE was increased in human HCC cell lines and tissues

The qRT-PCR results show that the expression level of IncRNA-CRNDE was significantly increased in seven HCC cell lines (MHCC97H, Hep3B, MHCC97L, BEL-7402, QGY-7703, HCCC9810, and HuH7) compared with immortalized normal liver epithelial cell lines (THLE3) (Figure 1A). Moreover, we detect the expression of IncRNA-CRNDE in 87 pairs of HCC tissues compared to paired adjacent normal tissues by qRT-PCR (Figure 1B). As shown in Table 1, the expression level (ΔCt) of IncRNA-CRNDE was related to the clinicopathological factors in HCC tissues. The ROC curve was generated, and the area under the ROC curve of IncRNA-CRNDE was 0.699 (P < 0.0001), which implies that it may serve as an ideal biomarker for HCC diagnosis (Figure 1C). Taken together, these results demonstrated that IncRNA-CRNDE was upregulated in HCC.

LncRNA-CRNDE promotes HCC cell proliferation, migration, and invasion in vitro

To investigate the biological function of IncRNA-CRNDE in the development process of HCC cells, we first detected the expression of IncRNA-CRNDE in MHCC97H cells transfected with IncRNA-CRNDE or the control by qRT-PCR. The results showed that IncRNA-CRNDE expression was significantly increased in MHCC97H cells transfected with IncRNA-CRNDE vector with lentivirus compared with the control (P < 0.001) (Figure 2A). Simultaneously, IncRNA-CRNDE expression was significantly decreased in HuH7 cells that were transfected with shRNA-CRNDE compared with the scramble (P < 0.001) (Figure 2B). Second, we measured the effect of IncRNA-CRNDE on the cell ability of HCC by MTT. As shown in Figure 2C, the overexpression of IncRNA-CRNDE promoted the proliferation of MHCC97H cells (P < 0.001). Meanwhile, the reduction of IncRNA-CRNDE markedly suppressed the proliferation of HuH7 cells (P...
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A

Relative IncRNA-CRNDE mRNA expression level (2^{-ΔΔCt})

MHCC97H Control
MHCC97H IncRNA-CRNDE

B

Relative IncRNA-CRNDE mRNA expression level (2^{-ΔΔCt})

HuH7 scramble
HuH7 shRNA

C

OD Value (A=490nm)

MHCC97H
MHCC97H vector
MHCC97H IncRNA-CRNDE

D

OD Value (A=490nm)

HuH7
HuH7 vector
HuH7 shRNA

E

MHCC97H

Control
LncRNA-CRNDE

Migration
Invasion

F

HuH7

Scramble
shRNA

Migration
Invasion

[Images and graphs depicting experiment results]
Figure 2. LncRNA-CRNDE promotes HCC cell proliferation, migration and invasion. A. The mRNA expression level of LncRNA-CRNDE was detected by qRT-PCR in MHCC97H cells transfected with LncRNA-CRNDE or the control (**p < 0.001). B. The mRNA expression level of LncRNA-CRNDE was detected by qRT-PCR in HuH7 cells transfected with shRNA-CRNDE or the scramble (**p < 0.001). C. The proliferation ability was measured by MTT assays in MHCC97H cells transfected with LncRNA-CRNDE or the control, (**p < 0.001). D. The silencing of LncRNA-CRNDE significantly inhibited cell proliferation. The proliferation ability was measured by MTT assays in HuH7 cells transfected with shRNA-CRNDE or the scramble (**p < 0.001). E. The overexpression of LncRNA-CRNDE significantly accelerated the migration and invasion ability of MHCC97H cells. Migrated and invasive cells were stained with crystal violet solution, and the quantification of migrated and invasive cells was shown; magnification 200 ×. ***p < 0.001. F. The silencing of LncRNA-CRNDE significantly inhibited migration and invasion ability of HuH7 cells. Migrated and invasive cells were stained with crystal violet solution, and the quantification of migrated and invasive cells was shown; magnification 200 ×, ***p < 0.001.

< 0.001) (Figure 2D). Finally, we detected the effect of LncRNA-CRNDE on the ability of HCC cell migration and invasion by transwell assays. As shown in Figure 2E, the number of migratory and invasive MHCC97H cells was significantly increased when cells were transfected with the overexpression vector of LncRNA-CRNDE (p < 0.001). In addition, the number of migratory and invasive HuH7 cells were significantly decreased when transfected with shLncRNA-CRNDE (p < 0.001) (Figure 2F).

**LncRNA-CRNDE negatively regulated miR-384 in human HCC**

The expression level of miR-384 was analyzed by qRT-PCR in 87 pairs of HCC tissues and paired adjacent normal tissues to determine if miR-384 is associated with the development process of HCC and LncRNA-CRNDE expression. The expression level of miR-384 was significantly lower in HCC tissues compared to paired adjacent normal tissues (p < 0.001) (Figure 3A). A correlation analysis found that there was a negative correlation between miR-384 and LncRNA-CRNDE mRNA by qRT-PCR in 87 pairs of HCC tissues and paired adjacent normal tissues (R² = 0.1335, P = 0.0005) (Figure 3B). Furthermore, we indicated that the expression level of miR-384 was significantly decreased in MHCC97H cells transfected with LncRNA-CRNDE compared with the control (p < 0.001), and the miR-384 expression was significantly increased when cells were transfected with miR-384 mimics again (p < 0.01) (Figure 3C). Simultaneously, we found that the expression level of miR-384 was significantly increased in HuH7 cells transfected with shRNA-CRNDE compared with the control (p < 0.01) and was significantly decreased when transfected with miR-384 inhibitors (p < 0.01) (Figure 3D). So, we further presumed that there is a binding site between miR-384 and LncRNA-CRNDE. StarBase v2.0 (http://starbase.sysu.edu.cn/) was used to predict the presumptive binding sites between LncRNA-CRNDE and miR-384. According to the results of the luciferase report assay, we showed that there was a significant decrease of the luciferase activities between the co-transfection of miR-384 mimics and wild-type LncRNA-CRNDE, with no changes in mutant LncRNA-CRNDE (Figure 3E). We also indicated that the proliferation ability was significantly increased in MHCC97H cells transfected with LncRNA-CRNDE compared with the control (p < 0.001), but it was significantly decreased in cells that were transfected with miR-384 mimics in combination (p < 0.001) (Figure 3F). Furthermore, we found that the proliferation ability of MHCC97H cells transfected with LncRNA-CRNDE was significantly decreased in HuH7 cells transfected with shRNA-CRNDE compared with the control (p < 0.001), yet, proliferation was significantly increased in cells transfected with miR-384 inhibitors in combination (p < 0.001) (Figure 3G). In short, these results showed that LncRNA-CRNDE promotes HCC cell proliferation though inhibition of miR-384.

**LncRNA-CRNDE regulates NF-κB and p-AKT expression**

In order to study the molecular mechanism LncRNA-CRNDE in HCC, qRT-PCR and Western blot experiments were used to detect the mRNA and protein expression levels of NF-κB and p-AKT in HCC cells. As shown in Figure 4A, the qRT-PCR results indicated that the mRNA expression level of NF-κB was significantly up-regulated in MHCC97H cells transfected with LncRNA-CRNDE compared with the control (p < 0.001) and was down-regulated when cells were transfected with miR-384 mimics in com-
LncRNA-CRNDE regulates miR-384 in HCC

A

Relative miR-384 mRNA expression level (ΔCT)

P < 0.0001

B

Expression level of lncRNA-CRNDE (ΔCT)

R² = 0.1335
P = 0.0005

C

MHCC97H

Relative miR-384 mRNA expression level

***

D

HuH7

Relative miR-384 mRNA expression level

***

E

LncRNA-CRNDE Wt 5′... UAAGGAUCUUUAACAAGUAU

Has-miR-384 3′... AUUCCUAGAAAUGUUGUCAU

LncRNA-CRNDE Mut 5′... UAAGGCCCUAGAUUAGUAU

F

Control

LncRNA-CRNDE

LncRNA-CRNDE+miR-384 mock

LncRNA-CRNDE+miR-384 mimics

G

Scramble

shRNA

shRNA+miR-384 mock

shRNA+miR-384 inhibitor
Figure 3. LncRNA-CRNDE negatively regulates miR-384 in human HCC. A. The qRT-PCR was used to detect the mRNA expression level of miR-384 in HCC tissues and paired adjacent normal tissues. The levels of transcript were normalized to U6 snRNA, (n = 87, ***P < 0.001). The correlation between LncRNA-CRNDE and miR-384 expression were measured (r = 0.546, P < 0.001). B. The expression level of miR-384 was measured by qRT-PCR in MHCC97H cells transfected with LncRNA-CRNDE or the control and in combination with miR-384 mock or mimics, (***P < 0.01, ***P < 0.001). C. The mRNA expression level of miR-384 was measured by qRT-PCR in MHCC97H cells transfected with LncRNA-CRNDE or the control, and in combination with miR-384 mock or inhibitors, (***P < 0.01). D. The expression level of miR-384 was measured by qRT-PCR in HuH7 cells transfected with shRNA-CRNDE or the scramble and in combination with miR-384 mock or inhibitors, (***P < 0.01). E. The correlation between LncRNA-CRNDE and miR-384 in the scramble and in combination with miR-384 mock or inhibitors, (***P < 0.01). F. The miR-384 inhibitors significantly decreased LncRNA-CRNDE-mediated cell proliferation. The proliferation ability was measured by MTT assays in MHCC97H cells transfected with lncRNA-CRNDE or the control, and then transfected with miR-384 mock or mimics, (***P < 0.001). G. The miR-384 inhibitors significantly promoted shRNA-CRNDE-mediated cell proliferation. The proliferation ability was measured by MTT assays in HuH7 cells transfected with shRNA-CRNDE or the scramble and then transfected with miR-384 mock or inhibitors (***P < 0.001).

Discussion

By reasons of poor prognosis, HCC is the most common type of liver cancer and is the major leading cause of cancer mortality all over the world [14, 15]. No completely effective treatment is available for primary liver cancer, however, the use of conventional radiotherapy and chemotherapy shows promise for prognosis [16, 17]. To date, studies indicate that several signaling pathways that are associated with biological functions participated in the development and progression of HCC [18, 19]. Particularly, the abnormally expressed proteins that are closely related to proliferation, migration, and invasion have been shown to be the main culprit in the formation process of liver cancer [20, 21]. Therefore, analyzing potential biomarkers is the most direct and effective way to explore the molecular mechanism and function for HCC.

Non-coding RNAs (ncRNAs) are segmented into two types, small ncRNAs and long non-coding RNAs (lncRNAs). As a class of small ncRNAs, miRNAs regulate the post-transcriptional level of target mRNAs resulting in mRNAs degradation [22, 23]. Some studies indicate that miR-122 [24], miR-372 [25], miR-375 [26] all participate in the process of liver cancer. LncRNA also has been shown to play important roles in gene regulation, such as epigenetic, transcriptional, posttranscriptional, and translational functions [27, 28]. Numerous studies have indicated that IncRNAs can act as competitive endogenous RNA (ceRNA) and play key roles in the occurrence and development of copious diseases [29, 30]. In this study, we found that the expression level of LncRNA-CRNDE was increased in human HCC. LncRNA-CRNDE promoted HCC cell proliferation, migration and invasion in vitro. Furthermore, we found that the upregulated LncRNA-CRNDE could act as a sponge for negatively regulated miR-384 and promoted HCC cells tumorigenesis.

NF-κB transcription factor, which acts as a regulator in cell survival, is involved in cancer development by encouraging a major inflammatory pathway activated during liver injury [31]. PI3K-Akt signaling pathway is regarded as a critical cause for tumorigenesis and plays major roles in cancer cell growth, survival, and proliferation. Members of the Akt pathway may become new targets for cancer treatment [32]. In our study, we first indicate that the LncRNA-CRNDE/miR-384 axis regulates these two important factors in HCC cell proliferation, invasion, and migration.
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A

MHCC97H

Relative NF-κB mRNA expression level (2ΔΔCt)

Control
LncRNA-CRNDE
LncRNA-CRNDE+miR-384 mock
LncRNA-CRNDE+miR-384 mimics

***

B

MHCC97H

Relative p-AKT mRNA expression level (2ΔΔCt)

Control
LncRNA-CRNDE
LncRNA-CRNDE+miR-384 mock
LncRNA-CRNDE+miR-384 mimics

***

C

MHCC97H

Control
LncRNA-CRNDE
LncRNA-CRNDE+miR-384 mock
LncRNA-CRNDE+miR-384 mimics

NF-κB
p-AKT
AKT
GAPDH

D

HuH7

Relative NF-κB mRNA expression level (2ΔΔCt)

Scramble
shRNA
shRNA+miR-384 mock
shRNA+miR-384 inhibitor

***

E

HuH7

Relative p-AKT mRNA expression level (2ΔΔCt)

Scramble
shRNA
shRNA+miR-384 mock
shRNA+miR-384 inhibitor

**

F

HuH7

Control
shRNA
shRNA+miR-384 mock
shRNA+miR-384 inhibitor

NF-κB
p-AKT
AKT
GAPDH

G

miR-384
LncRNA-CRNDE
NF-κB
P-AKT
Proliferation
Migration
Invasion

LncRNA-CRNDE
miR-384

Am J Cancer Res 2016;6(10):2299-2309
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Figure 4. LncRNA-CRNDE regulates NF-κB and p-AKT expression. (A) The mRNA expression level of NF-κB was measured by qRT-PCR in MHCC97H cells transfected with lncRNA-CRNDE or the control and then transfected with miR-384 mock or inhibitors. (**P < 0.001). (B) The mRNA expression level of p-AKT was measured by qRT-PCR in HuH7 cells treated as D (***P < 0.001). (C) The protein expression levels of NF-κB, AKT and p-AKT were detected by Western blotting in MHCC97H cells treated as A; GAPDH was used as a protein-loading control. (D) The mRNA expression level of NF-κB was measured by qRT-PCR in HuH7 cells transfected with shRNA-CRNDE or the scramble (**P < 0.01, ***P < 0.001). (E) The mRNA expression level of p-AKT was measured by qRT-PCR in HuH7 cells treated as D (**P < 0.01, ***P < 0.001). (F) The protein expression levels of NF-κB, AKT and p-AKT were detected by Western blotting in HuH7 cells treated as D; GAPDH was used as a protein-loading control. (G) A diagrammatic sketch of lncRNA-CRNDE functions in HCC. LncRNA-CRNDE promoted HCC cell proliferation, migration, and invasion though inhibition of miR-384.

In conclusion, this study can be summarized by following major findings: 1) lncRNA-CRNDE was unregulated in HCC tissues and cells and therefore has a positive correlation with clinicopathological features. 2) lncRNA-CRNDE was negatively correlated with miR-384 and accelerated the expression levels of NF-κB and p-AKT though inhibition of miR-384.

Acknowledgements

This study was Supported by the National key clinical specialist construction Programs of China, Nation Health Office Medical Case, No. (2013) 544.

Disclosure of conflict of interest

None.

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Am J Cancer Res 2016;6(10):2299-2309


